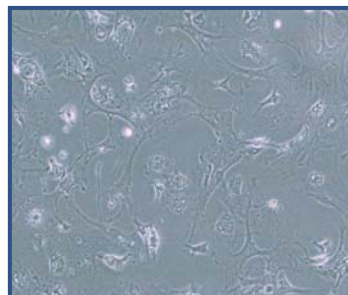


## MOUSE HEPATIC MYOFIBROBLASTS

### Introduction

Hepatic myofibroblasts (stellate cells) are isolated from 6-8 week-old ICR/CD-1 mice. Cell isolation has been performed aseptically by two-step collagenase perfusion of liver. An enriched fraction of hepatic myofibroblasts is obtained after separation from cell suspension in Percoll gradient. Primary cultured cells are allowed to grow for two weeks prior to cryopreservation.



### Safety Statements

Health quality controls regularly performed on these mice including bacteriology, virus serology and parasitology controls, certify these animals to be free of pathogens. Hepatic Myofibroblasts are PCR tested to be negative for Mycoplasma.

### Product Characteristics

**Organism:** *Mus musculus* (mouse)

**Tissue:** Liver.

**Cell type:** Hepatic Myofibroblasts (Stellate Cells)

**Mouse:** Male, ICR/CD-1.

**Number of viable cells post-thawing:**  $0,5 \times 10^6$  mouse hepatic myofibroblasts

**Purity:** > 99%

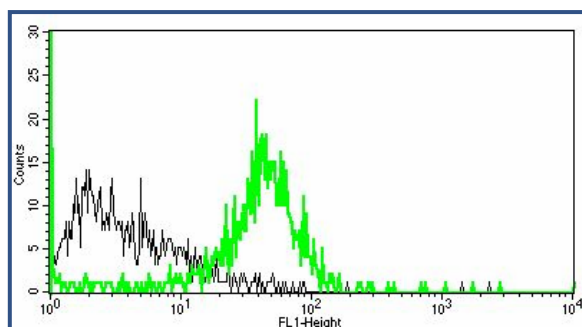
**High viability of cells,** measured by 7-amino-actinomycin viability probe (Via-Probe™), BD.

**Growth properties:** Adherent (plastic).

### Cell Characterization

Plated cells are visually checked for proper morphology.

Cells are characterised by flow cytometry for the phenotype-specific marker anti-Glial Fibrillary Acidic Protein (GFAP) shown in green. Unspecific isotype IgG2 is shown in black.



**Handling instructions:**

1. The recommended seeding density for attachment is approximately 7000 cells/cm<sup>2</sup> on plastic flasks/wells.
2. Use approximately 0.25 ml of culture medium per cm<sup>2</sup> of the plating surface and allow flasks/wells to equilibrate in a humidified 5% CO<sub>2</sub> incubator at 37°C for a minimum of 30 minutes.
3. Rapidly transfer the cryovial to thaw in a clean water bath at 37°C with gentle shaking. Monitor the contents of the vial, and when a small ice crystal remains remove the vial from the water bath. Wipe the cryovial with ethanol before opening.
4. Immediately mix the cell suspension using a micropipette avoiding the formation of bubbles and transfer it gently to the flasks previously prepared in step 2.
5. Help to distribute cells evenly by rocking the flasks gently. Loosen caps of flasks (unless vented caps are used) and return them to the incubator.
6. Change culture medium once cells are properly attached (no longer than 24 hours after seeding) and every second day thereafter. Pre-warm an appropriate amount of medium to 37°C before each medium change. Subculture cells when they are 70-90% confluent using Trypsin/EDTA solution.

\* Please, note that primary culture cells have a finite *in vitro* life-span.

**Culture medium**

Culture medium consists of RPMI (Sigma-Aldrich, R-8758) with 50 U/ml penicillin, 50 µg/ml streptomycin and 10% Fetal Calf Serum.

**Storage:**

Store in liquid nitrogen. For best results, use upon arrival.

Note:

- ▶ *Dominion Pharmakine SL guarantees the performance of this product only when the recommended storage and protocols are followed.*
- ▶ *If this product does not arrive in good condition, please contact your distributor*

